



Determination of Antioxidant Activity of Leave Extracts of *Albizia chevalieri* Using Free Radical Scavenging Activity Assay

Alhassan M. Garba^{1*}, Habiba R. Isa¹, Sadiq Abubakar² and Saudat Ja'afar¹

¹Department of Chemistry, Kano University of Science and Technology Wudil, P.M.B. 3244, Kano State, Nigeria.

²Department of Pure and Industrial Chemistry, Bayero University, Kano, P.M.B. 3011, Kano State, Nigeria.

Authors' contributions

This work was carried out in collaboration among all authors. Author AMG performed the statistical analysis, wrote the protocol and wrote the first draft of the manuscript. Author HRI designed the study. Author SA managed the analyses of the study and author SJ managed the literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IRJPAC/2019/v19i130099

Editor(s):

- (1) Dr. Hao-Yang Wang, Department of Analytical, Shanghai Institute of Organic Chemistry, Shanghai Mass Spectrometry Center, China.
- (2) Dr. Surendra Reddy Punganuru, Department of Biomedical Sciences, School of Pharmacy, Texas Tech University Health Sciences Center, Amarillo, USA.

Reviewers:

- (1) Eduardo Martinez-Abundis, Juarez Autonomous University of Tabasco, México.
- (2) Veeravan Lekskulchai, Srinakharinwirot University, Thailand.
- (3) Pratibha Kamble, Ohio State University, Columbus.

Complete Peer review History: <http://www.sdiarticle3.com/review-history/47673>

Original Research Article

Received 01 March 2019

Accepted 15 May 2019

Published 30 May 2019

ABSTRACT

Dried and powdered leaves of *Albizia chevalieri* were extracted using ethanol. The extract was fractionated to give methanol, chloroform and pet-ether. The four extracts obtained; ethanol, chloroform, methanol and pet-ether were evaluated for antioxidant activity using 2,2-diphenyl-1-(2,4,6-trinitrophenyl)hydrazyl (DPPH) free radical scavenging activity assay. The results of the DPPH scavenging activity indicated a concentration-dependent antioxidant activity. The DPPH scavenging activity of the ethanol, chloroform and methanol extracts were found to be promising. There is no significant difference in the antioxidant activity between the ethanol, chloroform and methanol extracts with that of standard Ascorbic acid at 10, 25, 250 and 500 µg/ml concentrations. This showed that the ethanol, chloroform and methanol leave extracts of the plant has the potency

*Corresponding author: E-mail: alhassangarba01@gmail.com, alhassan.garba01@gmail.com;

of scavenging free radicals *in vitro* and may provide leads in the ongoing search for natural antioxidants from Nigerian medicinal plants to be used in treating diseases related to free radical reactions.

Keywords: *Albizia chevalier*; antioxidant; free radical; extract; scavenging activity assay; DPPH.

1. INTRODUCTION

The plant *Albizia chevalieri* is a tree or a shrub that grows up to 12m height under harsh conditions of the dry savannah from Senegal, Niger and Nigeria. It has an open and rounded or umbrella-shaped canopy, bark pale-grayish, twigs pubescent with white lenticels, leaves with 8-12 pairs of pinnate and 20-40 pairs of leaflets each. The bark was reported to contain alkaloids and also tannin sufficient for use in tanning in Nigeria and Senegal. It is used in Borno-North eastern Nigeria as purgative, taenicide and also remedy for coughs. A decoction of leaves is used in Northern Nigeria as a remedy for dysentery [1]. There are also reports on the local use of the leaves extract for cancer treatment in Zaria city, Kaduna state [2].

Previous studies on *Albizia chevalieri* have indicated the presence of phenolic compounds from *Albizia amara* with significant antioxidant activity [3] and *Albizia inundata* was reported for effective anti candida activity from Brazilian flora [4]. Liphophilic extracts of *Albizia gummifera* revealed very promising antitrypanosomal activity [5]. The extracts of *Albizia ferruginea* were also reported to have significant antimicrobial activity on selected microorganisms [6] and *Albizia saman* was found to have good antiplasmodial activity [7]. *Albizia lebeck* was reported to contain 3 α , 5-dihydroxy-4 β , 7-dimethoxy flavones and N-Benzoyl-L-phenyl alaninol [8]. As the focus of medicine shifts from the treatment of manifest disease to prevention, increasing awareness on herbal remedies as potential sources of phenolic antioxidants have grown in recent years, and several plants are being screened for their antioxidant properties using different assays [9].

DPPH, known as 2,2-diphenyl-1-picrylhydrazyl, (I.U.P.A.C name, 2,2-diphenyl-1-(2,4,6-trinitrophenyl)hydrazyl with molecular formula C₁₈H₁₂N₅O₆ is a stable free radical that is commonly used to evaluate the ability of a compound to act as a free radical scavenger or hydrogen donor and to measure the antioxidant activity of tissue extract [10]. Free radicals are fundamental to any biochemical process and

represent an essential part of aerobic life and metabolism [11]. Antioxidants offer resistance against oxidative stress by scavenging the free radical and many other mechanisms thus preventing disease progression [12]. The reaction of DPPH with an antioxidant or reducing compound produces the corresponding hydrazine DPPH-H, which can be followed by color change from purple (absorbance at 515-528nm) to yellow. The DPPH method is widely used for the measurement of free radical scavenging ability of antioxidants [13,14]. DPPH is a rapid, simple, accurate and inexpensive assay for measuring the ability of different compounds to act as free radical scavengers or hydrogen donors and to evaluate the antioxidant activity of foods and beverages [15].

This work was designed to investigate the antioxidant properties of ethanol methanol, chloroform and pet-ether leaves extracts of *Albizia chevalieri* with a view to assessing the potential of the plant as a source for antioxidants.

2. MATERIALS AND METHODS

2.1 General

The Ethanol was obtained from Sigma Aldrich, the DPPH and Ascorbic acid was obtained from chemistry laboratory Bayero University Kano. While other reagents and chemicals were of analytical grade supplied by Chemistry Laboratory, Kano University of Science and Technology, Wudil. All glass wares used were washed with detergents and oven dried before use. The leaves of the plant *Albizia chevalieri* were rinsed with clean tap water to remove dust and impurities.

2.2 Collection of Plant Material

The leaves of plant *Albizia chevalieri* were collected on 4th November 2017 from Kududdufawa village Ungogo local government area of Kano State. The plant was authenticated by Baha'uddeen Said Adam from the Department of Plant Biology, Bayero University Kano, with accession number BUKHAN 0378.

as the radical is quenched by the antioxidant [19].

The leaves extracts of *Albizia chevalieri* were screened for DPPH radical scavenging activity according to the method described [20] with slight modification and the result of the screening is shown in (table 2) as compared to Ascorbic acid, a known antioxidant. Four different extracts of *Albizia chevalieri* (AC01, AC02, AC03, and AC04) showed high radical scavenging activity at various concentrations of 10, 25, 50, 100, 250 and 500 µg/ml (Fig. 2).

3.3 Inhibitory concentration at 50% (IC₅₀)

The radical scavenging activity of each extract was determined by calculating the inhibitory concentration at 50% (IC₅₀), the IC₅₀ of various extract of *Albizia chevalieri* and Ascorbic acid. The lower the IC₅₀, the more potent the extract, this showed that methanol extract has the highest radical scavenging activity (33.08 µg/ml) more than the standard Ascorbic acid (36.85 µg/ml), followed by ethanol extract (52.92 µg/ml), Chloroform extract (54.53 µg/ml) and pet-ether extract (71.59 µg/ml) [Fig. 3].

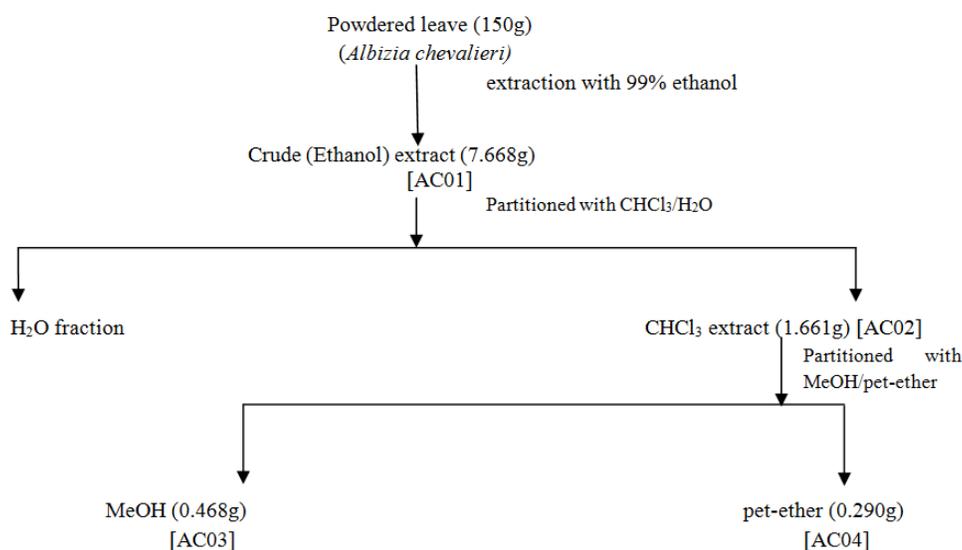


Fig. 1. Extraction and fractionation procedure of the powdered leaf of the plant

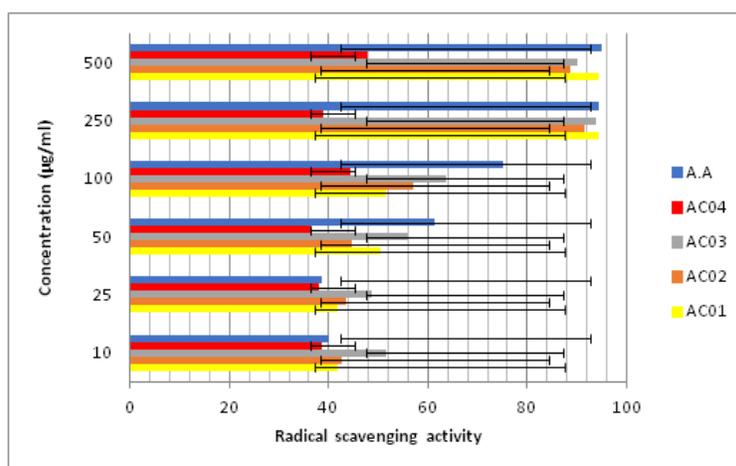
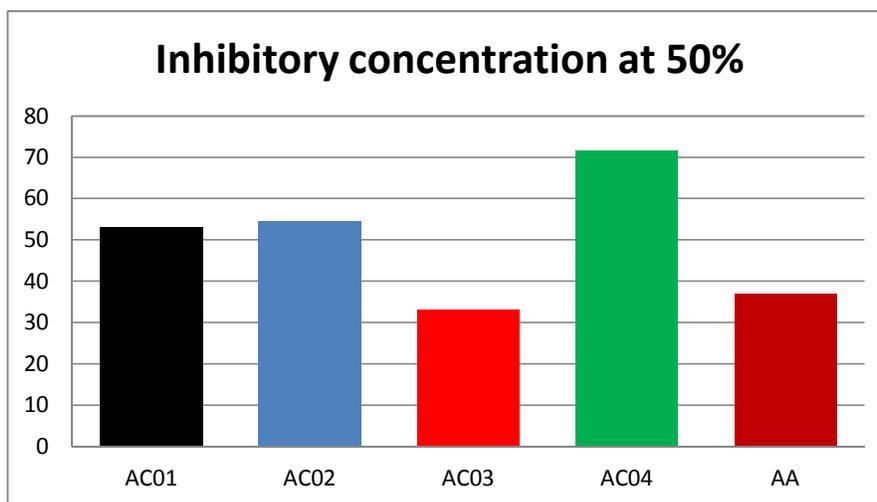


Fig. 2. Free radical scavenging activity of various extract of *Albizia chevalieri* at different concentrations

Table 1. Weights of extracts recovered and their physical properties

Extracts	Colour	Texture	Weight (g)	Weight (%)
AC01 [crude extract]	Dark green	Gummy like	7.668	5.11
AC02 [CHCl ₃ extract]	Black	Semi-solid	1.661	1.11
AC03 [methanolic extract]	Black	Semi-solid	0.468	0.31
AC04 [pet-ether extract]	Black	Semi-solid	0.290	0.19

**Fig. 3. Inhibitory concentration at 50% (IC₅₀) of various extracts of *Albizia chevalieri***

4. CONCLUSION

These findings revealed the potential of *Albizia chevalieri* as a source for natural antioxidants. It indicates that the plant could be a promising agent in scavenging free radicals and treating diseases related to free radical reactions. The leaves extracts of *Albizia chevalieri* were found to have high radical scavenging activity as compared with standard Ascorbic acid. The results of the DPPH scavenging activity study indicate a concentration-dependent antioxidant activity which increases with increase in the concentration of the extract.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Burkill HM. The useful plant of west tropical Africa. Families J-L. Royal Botanical Gardens, Kew. 1995;3:207–208.
- Aliyu AB, Musa AM, Ibrahim MA, Ibrahim H, Oyewale AO. Preliminary phytochemical screening and antioxidant activity of leave extract of *Albizia chevalieri* harms. Bayero Journal of Pure and Applied Sciences. 2009;1(2):149-153.
- Muchuweti M, Nyamukonda L, Chagonda, LS, Ndhala AR, Mupure C, Behura M. Total phenolic content and antioxidant activity in selected medicinal plants of Zimbabwe. Int. J. Food Sci. Tech. 2006;41: 33–38.
- Tempone AG, Sartorelli P, Teixeira D, Prado FO, Calixto IARL, Lorenzi H, Melhem M. Brazilian flora extract as source of nivel anileishmanial and antifungal compound. Mem, 1st Oswaldo Cruz. 2008;103(5):443–449.
- Freiburghaus F, Omgwal EN, Nkuny MH, Kaminsky R, Brun R. *In vitro* anti-trypanosomal activity of African plants used in traditional medicine in Uganda to treat sleeping sickness. Trop. Med, Int. Health. 2007;1(6):765–771.
- Agyare C, Kofuer GA, Mensah AY, Agyemang DO. Boletin Latinoamericano Y. del Criba de plantas Medicinales. Y Aromatica. 2006;5(2):31–35.
- Kohlera I, Jenett-Siema K, Siemsb K, Herna ndezc MA, Ibarrac RA,

- Berendsohnd WG, Bienzlee U, Eicha E. *In vitro* antiplasmodial investigation of medicinal plants from El Salvador. *Naturforsch.* 2002;57c:277–281.
8. Rashid RB, Chowdhury R, Jabbar A, Hassan GM, Rashid MA. Constituents of Albizia lebbeck and antibacterial activity of isolated flavones derivatives. *Saudi Pharm. J.* 2003;11(1-2):52–56.
 9. Karou D, Ndaembega WMC, Outtara L, Ilboudo DP, Canini A, Nikiema JP, Sempore J, Collizi V, Traore AS. African ethnopharmacology and new drug discovery. *Medicinal and Aromatic plant Science and Biotechnology.* 2006;1:1–9.
 10. Kedare Sagar B, Singh RP. Genesis and development of DPPH method of antioxidant assay. *J Food Sci Technol.* 2011; 48(4):412-422.
 11. Tiwari A. Imbalance in antioxidant defense and human diseases: multiple approach of natural antioxidants therapy. *Curr. Sci.* 2001;81:1179-1187.
 12. Brauggler JM, Duncan CA, Chase LR. The involvement of Iron in lipid peroxidation: Importance of ferrous to ferric ratio in initiation. *J. Biol. Chem.* 1986;61: 102-182.
 13. Perez–Jimenez J, Saura–Calixto F. Antioxidant capacity of dietary polyphenols determined by ABTS assay: A kinetic expression of the results”. *International Journal of food science and Technology.* 2008;43:185-191.
 14. Perez–Jimenez J, Arranz S, Tabernero M, Diaz-Rubia ME, Serrano J, Goni I, Saura-Calixto F. Updated methodology to determine antioxidant capacity in plant foods, oils and beverages: Extraction measurement and expression of results). *Food Research International.* 2008;41(3): 274–285.
 15. Prakash A. Antioxidant activity medallion laboratories analytical progress. 2001; 19(2).
 16. Fatope MO, Ibrahim H, Takeda Y. Screening of higher plants reputed as pesticides using brine shrimp lethality assay. *Int. J. Pharmacog.* 1993;31:250-56.
 17. Bozin B, Mimica-Dukic N, Samojlik I, Goran A, Igic R. Phenolics as antioxidants in garlic (*Allium sativum* L., Alliaceae). *Food Chem.* 2008;111:925-929.
 18. Huang D, Ou B, Prior RL. The chemistry behind antioxidant capacity assays. *J. Agric. Food Chem.* 2005;53:1841-1856.
 19. Karagozler AA, Erdag B, Emek YC, Uygum DA. Antioxidant activity and proline content of leaf extracts from *Dorystoechas hastate*. *Food Chem.* 2008;111:400-407.
 20. Mensor LI, Menezes FS, Leitao GG, Reis AS, dos Santos T, Coube CS, Leitao SG. Screening of Brazilian plants extracts for antioxidant activity by the use of DPPH free radical method. *Phytother. Res. Tech.* 2001;15:127–130.

© 2019 Garba et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:

<http://www.sdiarticle3.com/review-history/47673>