

The Importance of Increased Serum Ornithine Levels in the Pathogenesis of Alzheimer and Parkinson's Diseases

V. Kenan Çelik^{1*}, Burhanettin Çiğdem², Serkan Kapancik¹, Hasan Kiliçgün³ and Ertuğrul Bolayir²

¹Department of Biochemistry, Faculty of Medicine, Cumhuriyet University, Sivas, Turkey.

²Department of Neurology, Faculty of Medicine, Cumhuriyet University, Sivas, Turkey.

³Department of Nutrition and Dietetics, Faculty of Health Science, University of Erzincan, Turkey.

Authors' contributions

This work was carried out in collaboration between all authors. Authors VKC and EB designed and supervised the study. Author VKC wrote the protocol and wrote the first draft of the manuscript. Author BC collected of patient and control serum. Authors BC and SK managed the analyses of the study. Author SK managed the literature searches. Author HK performed the statistical analysis and interpretation. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/AJORRIN/2018/42254

Editor(s):

(1) Dr. Ekanem Eyo Philip-Ephraim, Associate Professor, Department of Internal Medicine, University of Calabarteaching Hospital, Nigeria.

Reviewers:

(1) T. Hiwasa, Chiba University, Japan.

(2) Maria Rosa Avila-Costa, National Autonomous University of Mexico, Mexico.

(3) Hanaa Hamdy Ahmed, Egypt.

Complete Peer review History: <http://www.sciencedomain.org/review-history/25606>

Original Research Article

Received 3rd May 2018
Accepted 11th July 2018
Published 19th July 2018

ABSTRACT

Background: The aim of this study was to investigate the levels of enzymes and ornithine involved in the synthesis of polyamines in patients with Alzheimer's and Parkinson's disease and to see their positive or negative effects on the modulation of the immune system.

Methods: Thirty-five healthy subjects as a control group and 35 patients with Alzheimer's and Parkinson's disease were included in this study. Determination of Ornithine decarboxylase, Arginine Decarboxylase and Agmatinase levels were evaluated by using Enzyme-Linked Immunosorbent Assay (Elisa kit). Ornithine levels were measured spectrophotometrically.

Results: When ornithine levels of Alzheimer and Parkinson patients were compared to the control

group, differences were found as significant ($p < 0.05$). On the other hand, when Ornithine decarboxylase, Arginine Decarboxylase, Agmatinase levels of Alzheimer and Parkinson patients were compared to the control group, the differences were found as insignificant ($p > 0.05$).

Conclusion: Although the enzyme levels in the pathway of polyamine synthesis in Alzheimer's and Parkinson's diseases do not change, the increase in ornithine level will not contribute to the fight mode of the immune system. On the contrary, it may change plasticity by increasing osmolality.

Keywords: Alzheimer; Parkinson; polyamine; ornithine; agmatine; immunomodulation.

1. INTRODUCTION

Alzheimer's and Parkinson's disease are two of the most common neurological diseases that cause neurodegeneration and affect many people [1]. The most valid hypothesis explaining the development of Alzheimer's disease (AD), the most common neurological disease, is the amyloid cascade hypothesis. According to this hypothesis, amyloid- β peptides ($A\beta$) accumulate in the cerebral blood vessels and brain parenchyma, and accumulation of hyperphosphorylated tau proteins in neurons leads to lethal disease, progressive loss of consciousness, functional impairment and memory loss [2,3]. Parkinson's disease, which is the second most common neurological disease that occurs after the death of dopaminergic neurons in Substantia nigra pars compacta, causes rigidity, tremor, and hypokinesia [4,5]. In addition, α -synuclein protein, which accumulates in neurons, plays a key role in Parkinson's disease. Lewy particles, which are caused by excessive accumulation of this protein in neurons, leading to pathology leading to disease progression in cholinergic and monoaminergic neurons in the brain. The diagnosis of idiopathic Parkinson's disease can be determined by applying these two major neuropathologies (neuronal loss in specific areas of the substantia nigra and widespread intracellular protein (α -synuclein) accumulation [6]. In Alzheimer's disease, a different pattern of α -synuclein pathology was found to accumulate mainly in the limbic region of the brain [7]. In vitro experiments have shown that the presence of polyamines in α -synuclein accumulation and fibril formation is effective and this effective sequence has been shown as spermin>spermidine>putrescine [8]. Although the mechanism of brain atrophy and neuronal loss is not fully known, there is a growing body of evidence recently that the lack of arginine and suppressing immunity plays a critical role in the pathogenesis of AD [9]. The cells responsible primarily for the immune system in the brain are microglia cells and macrophage-like immune cells found in the brain parenchyma,

which are involved in the modulation of the brain's inflammatory response [10]. The microglia are activated immediately after the ischemia. Circulating monocytes are rapidly transformed into macrophages in the brain via the blood-brain barrier due to the inflammation [10,11]. Microglia and macrophages are known to be essential cells in inflammation after cerebral ischemia. Polyamines have a negative regulatory effect on macrophage activation through complex associations with NO metabolism. In macrophages, NO is an intermediate product in the L-arginine oxidation process [12].

Polyamines are molecules having 2, 3 or 4 amino groups. They are widely found in living organisms because they have a key role in the survival of life [13]. The major polyamines synthesized by several enzymes from the amino acid of L-Arginine are putrescine, spermidine, spermine and agmatine (Fig. 1). Polyamines play an important role in biological processes because of their interaction with many different receptors, protein kinases, nucleotide cyclases. It has been reported that the increase in the levels of polyamines may be associated with diseases such as cancer, as they have a key role in growing and dividing the cell [14,15,16]. However, due to their interaction with systems such as polyamines, catecholamines, GABA, nitric oxide (NO), glutamate, they are also associated with many psychiatric disorders, especially schizophrenia [16,17,18]. The survival of the cells that make up the organism depends on the presence of polyamines. It is known that neurodegenerative diseases caused by cell death develop due to the differentiation of polyamine metabolism [19,20,21]. Many researchers have shown that agmatine has neuroprotective potential and develops a cognitive function in various animal models of central nervous system damage such as neurotrauma and neonatal ischemia [22,23,24,25,26]. The degraded polyamine metabolism indirectly causes the degradation of nitric oxide synthase. Increased agmatine synthesis causes suppression of NO synthesis in

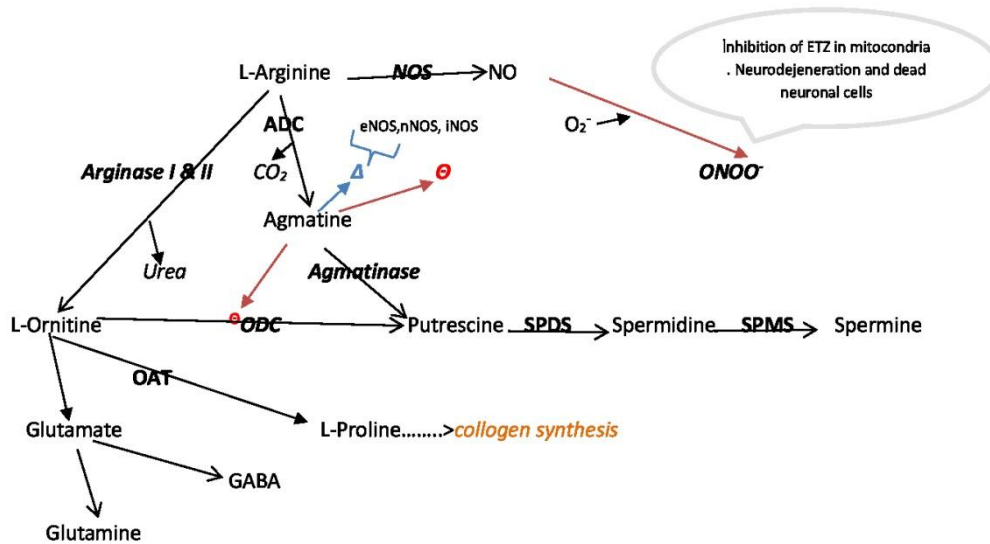


Figure 1: Regulation of NO synthesis and polyamines synthesis by agmatine. Nitric oxide synthase (NOS); endotelialNOS(eNOS); neural NOS(nNOS); inducible NOS (iNOS); Arginine decarboxylase (ADC); Ornithine decarboxylase (ODC); Spermidine synthase SPDS; Spermine synthase SPMS; γ -aminobutyric acid (GABA); Ornithine amino tranferase (OAT); Ornithine decarboxylase (ODC); inhibition; \ominus ; Activation Δ :

immunocytes and leads to defense deficiency, while decreased levels result in increased NO synthesis resulting in neurodegeneration and death of neurons (Fig. 1) [27,28,29].

Our aim in this study was to reveal the possible roles of polyamines in the pathogenesis of Alzheimer and Parkinson diseases. For this purpose, the levels and effects of arginine decarboxylase, ornithine decarboxylase, agmatinase and ornithine involved in the pathway of polyamine synthesis on the immune system were examined.

2. MATERIALS AND METHODS

2.1 Patient and Control Group

The 35 patients, who applied to the Cumhuriyet University Medicine Faculty Neurology Polyclinic and took the diagnosis of Alzheimer's and Parkinson, were included in the study as an experimental group. Patients of all ages and gender were included in the study. Our control group was chosen from the 35 healthy individuals who have not any systemic diseases (diabetes, hypertension and neurodegenerative disease).

2.2 Blood Samples Collection

Blood samples were taken from the controls and patients who were made the diagnosis of

Alzheimer and Parkinson and not receive any therapy. The serums were obtained by centrifugation at 1610 x g for 10 minutes of the blood samples were preserved in 80 0C to study.

2.3 Detection of Enzyme Levels

Ornithine, arginine decarboxylase, ornithine decarboxylase and agmatinase levels have been measured in serum of Alzheimer and Parkinson patients. Ornithine level was calculated spectrophotometrically at 515 nm using the method defined by Chinard and the value was given as $\mu\text{mol/ml}$ [30]. Arginine decarboxylase, ornithine decarboxylase, and agmatinase levels were determined by ELISA kit according to the manufacturer's protocol (SunRed, China).

2.4 Statistical Analysis

Mann Whitney U test was used for statistical analysis of the data. The data were stated in the tables as the arithmetical average \pm Standard deviation and the level of significance was taken as 0,05.

3. RESULTS

There wasn't any significant difference between patient groups and the control group in terms of gender and age ranges ($P > 0.05$) (Table 1).

When ADC, ODC and Agmatinase enzymes, which were determined at serum levels, were compared according to the control group, no statistically significant difference was found between the two disease groups (Alzheimer and Parkinson) ($P>0.05$) (Table 2).

On the other hand, serum ornithine levels increased in both groups when compared with the control group, and they were statistically significant. $P<0.05$ (Table 2).

4. DISCUSSION

In this study, enzymes (ADC, ODC, and Agmatinase) and ornithine levels in polyamine synthesis pathway were measured in serums of Alzheimer and Parkinson's patients. When the levels of the enzymes were compared with the control as a result of the analysis, no significant difference was found between the two groups of patients and control ($p>0.05$). Ornithine was higher and statistically significant in both groups when compared to the control group ($p<0.05$). Enzyme and ornithine levels in the polyamine synthesis pathway have been investigated in this study for the first time in serums of Alzheimer and Parkinson's patients. Most of the studies carried out to date regarding polyamines and

their relation to these diseases were in the form of experimental animals and postmortem studies. In one postmortem study, the amount of spermidine increased in the temporal cortex (70%) and a reduction in levels of putrescine (28%) was found in patients with AD [19]. In another postmortem study, there was no difference in spermidine and spermine concentrations in the basal ganglia of Parkinson, Huntington's disease (HD) and progressive supranuclear palsy (PSP) patients and it was found to decrease with age [31]. Changes in the homeostasis of polyamines play an important role in the emergence of many diseases such as cell growth, senility, memory performance, neurodegenerative diseases, metabolic diseases and cancer [32,33].

In the polyamine synthesis pathway, two major molecules play an important role in the modulation of the immune system. These are ornithine and agmatine molecules. Ornithine is the precursor molecule of the synthesis of major polyamines (putrescine, spermidine and spermine), the precursor of both L-glutamate and GABA molecules as well as the precursor of L-proline required for the synthesis of connective tissue for wound healing (Figure 1). In this study, ornithine levels were increased in both groups of

Table 1. The gender and age ranges of patients and control

Gender	Control	Alzheimer patient	Control	Parkinson Patient
Male	25	25	19	19
Women	10	10	16	16
Age (X ± S)	77 ± 11	74 ± 14	67 ± 12	68 ± 11

Data expressed as mean ± standard deviation

Table 2. The serum ornithine, arginine decarboxylase, ornithine decarboxylase and agmatinase levels in patients and controls

	Alzheimer			Parkinson		
	Control	Patient	p	Control	Patient	p
ADC (pg/ml) (n=35)	3.07±0.22	3.08±0.13	>0.05	3.11± 0.21	3.07±0.22	>0.05
Agmatinase (pg/ml) (n=35)	3.06±0.2	3.02±0.21	>0.05	3.09±0.21	3.06±0.13	>0.05
ODC (pg/ml) (n=35)	3.13±0.15	3.11±0.23	>0.05	3.11±0.21	3.07±0.2	>0.05
Ornithine (µmol/ml) (n=35)	0.12±0.02	0.16±0.02	<0.05	0.14±0.03	0.16±0.02	<0.05

patients compared to the control and statistically significant ($p < 0.05$). The major metabolic ways in which the increased ornithine can go, are L-proline, L-glutamate, GABA, polyamine synthesis and the urea cycle. Since the ODC levels, the rate-limiting enzyme in polyamine synthesis did not change in both patient groups, the polyamine synthase pathway would not be active (Table 2).

The entry of the increased ornithine into the urea cycle will contribute to the formation of excessive urea and thus the increase of osmolarity in many regions of the brain such as the cerebellum, cerebral cortex and brain stem [34]. Chronic osmotic pressure caused by ornithine also modifies plasticity in the hippocampal region by mediating tonic inhibition of the GABA receptor family [18,35,36]. On the other hand, the increase of L-proline synthesis for collagen production, which is necessary for connective tissue, means that immune system cells pass into the fixed mode (wound healing). Thus, the increased L-proline synthesis will only contribute to the synthesis of collagen [37]. The main problems of both Parkinson and Alzheimer's patients are mainly a cluster of α -synuclein and amyloid beta. Extra collagen accumulation will also increase the negative effect. This approach could not be verified for the time being because L-proline levels could not be measured due to the limitations of the study. Another molecule that is effective in the modulation of immune cells is agmatine. The active immunocytes (microglia, macrophage) increase NO production in the M1 (fight-killing program) mode. They do this via inducible nitric oxide synthase (iNOS). Nitric oxide is a twisted sharp knife; The first is the main events caused by their decreasing concentration. These are reduced vascular perfusion, blockage of granulocytes in the blood vessels in the inflamed area, inhibition of pro-inflammatory reactions, and thickening of the capillary membranes [38,39,40]. All these adverse events lead to neurodegeneration and neuronal cell death due to inadequate oxygen and glucose transport to the resulting neuronal and glial cells [41]. In the second, high concentrations of NO inhibit many complexes in the respiratory chain and lead to an increase in the synthesis of highly toxic compound peroxynitrite (ONOO^-). This highly toxic compound causes mitochondrial damage and leads to mitochondrial dysfunction leading to neurodegeneration and neuronal cell death [42]. When the level of arginine is reduced to an undetectable level in regions with inflammation, it can be understood that Arginine metabolism is a

central pathway for macrophages in the immune system [43]. When inflammation occurs, the macrophages must be directed to one of the "fight" or "fix" modes. The fight is a killing program (eg pathogen) and fix is a wound healing program. The Fight pathway activated by macrophages results in the formation of nitric oxide (NO) and cell proliferation is inhibited [44]. When the fixed pathway is activated, proliferation and healing process is initiated by ornithine formation (via polyamine and collagen) [45,46,47,48]. Both the M1 (fight) and M2 (fix) pathways use arginine, but the NO and Ornithine pathways cannot be active at the same time because the metabolites of a pathway inhibit the enzymes of the other pathway (Şekil 1). The fact that ADC enzyme levels which are effective in the synthesis of agmatine in both groups of patients do not change and that there is no difference in agmatinase enzyme in the destruction pathway will not change the amount of agmatine (Table 2). Solubulous amyloid-Beta oligomers are responsible for synapse loss and neurodegenerative development in Alzheimer's disease [49]. B-Amyloid 1-40 peptide regulates glutamate release by acting on glutamatergic terminals [50], while B-amyloid plaques lead to increased intracellular calcium concentration and death of neurons by directly affecting NMDA receptors [51].

The absence of an increase in the amount of agmatine, the natural antagonist of the receptors, naturally inhibits blockage of these receptors.

Agmatine activates eNOS while inhibiting iNOS from NOS enzymes. Studies on experimental animal models showed that application of agmatine in brain cells damaged by lipopolysaccharide reduced and corrected the damage with these properties [52,53,54,55].

5. CONCLUSION

As a conclusion, polyamine synthesis enzymes whose levels are not altered in Alzheimer and Parkinson's disease cannot contribute to the prevention of neurodegeneration. Increased ornithine levels will not contribute to the fight mode of the immune system but may contribute to fixing mode which increased proline's levels. But, it may contribute to the synthesis and increase of urea and impair plasticity. Also, it may assist to glutamate, GABA and urea synthesis and increase osmolarity and impair plasticity. Exposing these grey areas with more extensive studies may allow new approaches

to be introduced in the treatment of such diseases.

CONSENT

As per international standard or university standard written participant consent has been collected and preserved by the authors.

ETHICAL APPROVAL

As per international standard or university standard written ethical permission has been collected and preserved by the authors.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Lleó A, Cavedo E, Parnetti L, Vanderstichele H, Herukka SK, Andreasen N, Ghidoni R, Lewczuk P, Jeromin A, Winblad B, Tsolaki M, Mroczko B, Visser PJ, Santana I, Svenningsson P, Blennow K, Aarsland D, Molinuevo JL, Zetterberg H, Mollenhauer B. Cerebrospinal fluid biomarkers in trials for Alzheimer and Parkinson diseases. *Nature Reviews Neurology*. 2015;11(1):41-55.
- Querfurth HW, LaFerla FM. Alzheimer's disease. *N. Engl. J. Med.* 2010;362:329–344.
- Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. *Science*. 2002;297:353–356.
- Conway KA, Harper JD, Lansbury PT. Lansbury accelerated in vitro fibril formation by a mutant alpha-synuclein linked to early-onset Parkinson disease. *Nat Med*. 1998;4:1318–1320.
- Klein C, Schlossmacher MG. Parkinson disease, 10 years after its genetic revolution: Multiple clues to a complex disorder. *Neurology*. 2007;69(22):2093-2104.
- Poewe W, Seppi K, Tanner CM, Halliday GM, Brundin P, Volkman J, Schrag AE, Lang AE. Parkinson disease. *Nature Reviews Disease Primers*. 2017;3:17013.
- Iacono D, Geraci-Erck M, Rabin ML, Adler CH, Serrano G, Beach TG, Kurlan R. Parkinson disease and incidental Lewy body disease Just a question of time? *Neurology*. 2015;85(19):1670-1679.
- Antony T, Hoyer W, Cherny D, Heim G, Jovin TM, Subramaniam V. Cellular polyamines promote the aggregation of α -synuclein. *Journal of Biological Chemistry*. 2003;278(5):3235-3240.
- Kan MJ, Lee JE, Wilson JG, Everhart AL, Brown CM, Hoofnagle AN, Jansen M, Vitek MP, Gunn MD, Colton CA. Arginine deprivation and immune suppression in a mouse model of Alzheimer's disease. *Journal of Neuroscience*. 2015;35(15): 5969-5982.
- Ginhoux F, Lim S, Hoeffel G, Low D, Huber T. Origin and differentiation of microglia. *Front Cell Neurosci*. 2013;7:45. DOI: 10.3389/fncel.2013.00045
- Bonaventura A, Liberale L, Vecchié A, Casula M, Carbone F, Dallegri F, Montecucco F. Update on inflammatory biomarkers and treatments in ischemic stroke. *Int J Mol Sci*. 2016;17(12):E1967. DOI: 10.3390/ijms17121967
- Marletta MA, Yoon PS, Iyengar R, Leaf CD, Wishnok JS. Macrophage oxidation of L-arginine to nitrite and nitrate: Nitric oxide is an intermediate. *Biochemistry*. 1988;27: 8706–8711.
- Tabor CW, Tabor H. Polyamines. *Annu Rev Biochem*. 1984;53:749-790.
- Bachrach U. Polyamines and cancer: Minireview article. *Amino Acids*. 2004;26: 307-309.
- Kapancik S, Celik VK, Kilickap S, Kacan T, Kapancik S. The relationship of agmatine deficiency with the lung cancer. *International Journal of Hematology and Oncology*. 2016;26(4):103-109.
- Piletz JE, Aricioglu F, Cheng JT, Fairbanks CA, Gilad VH, Haenisch B, Halaris A, Hong S, LeeJE, Li J, Liu P, Molderings GJ, Rodrigues ALS, Satriano J, Seong GJ, Wilcox G, Wu N, Gilad GM. Agmatine: Clinical applications after 100 years in translation. *Drug Discovery Today*. 2013;18(17):880-893.
- Uzbay T. An alternative approach to understand schizophrenia: Polyamine hypothesis through NMDA receptors; 2014.
- Çelik VK, Ersan EE, Kilicgun H, Kapancik S, Ersan S. Agmatine mediated hypertonic stress development in schizophrenia: A novel study. *Neuropsychiatry*. 2016;6(5).
- Morrison LD, Kish SJ. Brain polyamine levels are altered in Alzheimer's disease. *Neuroscience Letters*. 1995;197(1):5-8.

20. Minois N, Carmona-Gutierrez D, Madeo F. Polyamines in aging and disease. *Aging (Albany NY)*. 2011;3(8):716-732.
21. Lewandowski NM, Ju S, Verbitsky M, Ross B, Geddie ML, Rockenstein E, Adame A, Muhammad A, Vonsattel JP, Ringe D, Cote L, Lindquist S, Masliah E, Petsko GA, Marder K, Clark LN, Small SA. Polyamine pathway contributes to the pathogenesis of Parkinson disease. *Proceedings of the National Academy of Sciences*. 2010; 107(39):16970-16975.
22. Liu P, Collie ND. Behavioral effects of agmatine in naive rats are task- and delay-dependent. *Neuroscience*. 2009;163:82-96.
23. Lu W, Dong HJ, Gong ZH, Su RB, Li J. Agmatine inhibits morphine-induced memory impairment in the mouse step-down inhibitory avoidance task. *Pharmacol Biochem Behav*. 2010;97:256-61.
24. McKay BE, Lado WE, Martin LJ, Galic MA, Fournier NM. Learning and memory in agmatine-treated rats. *Pharmacol Biochem Behav*. 2002;72:551-7.
25. Zarifkar A, Choopani S, Ghasemi R, Naghdi N, Maghsoudi AH, Maghsoudi N, Rastegara K, Moosaviat M. Agmatine prevents LPS-induced spatial memory impairment and hippocampal apoptosis. *Eur J Pharmacol*. 2010;634:84-8.
26. Kim JH, Yenari MA, Giffard RG, Cho SW, Park KA, Lee JE. Agmatine reduces infarct area in a mouse model of transient focal cerebral ischemia and protects cultured neurons from ischemia-like injury. *Exp Neurol*. 2004;189:122-30.
27. Feng Y, Piletz JE, Leblanc MH. Agmatine suppresses nitric oxide production and attenuates hypoxic-ischemic brain injury in neonatal rats. *Pediatr Res*. 2002;52:606-11.
28. Takashi T, Katsuse O, Iseki E. Nitric oxide pathways in Alzheimer's disease and other neurodegenerative dementias. *Neurological Research*. 2004;26(5):563-566.
29. Gatto EM, Riobó NA, Carreras MC, Cherňavsky A, Rubio A, Satz ML, Poderoso JJ. Overexpression of neutrophil neuronal nitric oxide synthase in Parkinson's disease. *Nitric Oxide*. 2000;4(5):534-539.
30. Chinard FP. Photometric estimation of proline and ornithine. *Journal of Biological Chemistry*. 1952;199:91-95.
31. Vivó M, de Vera N, Cortés R, Mengod G, Camón L, Martínez E. Polyamines in the basal ganglia of human brain. Influence of aging and degenerative movement disorders. *Neuroscience Letters*. 2001; 304(1):107-111.
32. Miller-Fleming L, Olin-Sandoval V, Campbell K, Ralser M. Remaining mysteries of molecular biology: The role of polyamines in the cell. *Journal of Molecular Biology*. 2015;427(21):3389-3406.
33. Çelik VK, Kapancık S, Kaçan T, Kaçan SB, Kapancık S, Kılıçgün H. Serum levels of polyamine synthesis enzymes increase in diabetic patients with breast cancer. *Endocrine Connections*. 2017;6(8):574-579.
34. Sadasivudu B, Rao TI. Studies on functional and metabolic role of urea cycle intermediates in brain. *J. Neurochem*. 1976;27(3):785-794.
35. Pocklington AJ, Rees E, Walters JT, Han J, Kavanagh DH, Chambert KD, Holmans P, Moran JL, McCarroll SA, Kirov G, O'Donovan MC, Owen MJ. Novel findings from cnvs implicate inhibitory and excitatory signaling complexes in schizophrenia. *Neuron*. 2015;86(5):1203-1214.
36. Glykys J, Mann EO, Mody I. Which GABAA receptor subunits are necessary for tonic inhibition in the hippocampus? *J. Neurosci*. 2008;28(6):1421-1426.
37. Ginguay A, Cynober L, Curis E, Nicolis I. Ornithine aminotransferase, an important glutamate-metabolizing enzyme at the crossroads of multiple metabolic pathways. *Biology*. 2017;6(1):18.
38. Buée L, Hof PR, Bouras C, Delacourte A, Perl DP, Morrison JH, Fillit HM. Pathological alterations of the cerebral microvasculature in Alzheimer's disease and related dementing disorders. *Acta Neuropathol*. 1994;87:469-480.
39. de la Torre JC, Mussivand T. Can disturbed brain microcirculation cause Alzheimer's disease? *Neurol Res*. 1993;15:146-153.
40. Mancardi GL, Perdelli F, Rivano C, Leonardi A, Bugiani O. Thickening of the basement membrane of cortical capillaries in Alzheimer's disease. *Acta Neuropathol*. 1980;49:79-83.
41. de la Torre JC, Stefano GB. Evidence that Alzheimer's disease is a microvascular disorder: The role of constitutive nitric oxide. *Brain Res Brain Res Rev*. 2000;34: 119-136.

42. Stewart VC, Heales SJ. Nitric oxide-induced mitochondrial dysfunction: Implications for neurodegeneration. *Free Radic Biol Med.* 2003;34:287–303.
43. Albina JE, Mills CD, Henry WL Jr, Caldwell MD. Temporal expression of different pathways of L-arginine metabolism in healing wounds. *J Immunol.* 1990; 144(3):877–80.
44. Hibbs JB, Vavrin Z, Taintor RR. L-arginine is required for expression of the activated macrophage effector mechanism causing selective metabolic inhibition in target cells. *J Immunol.* 1987;138:550–65.
45. Mills CD. Macrophage arginine metabolism to ornithine/urea or nitric oxide/citrulline: A life or death issue. *Crit Rev Immunol.* 2001;21:399–425.
46. Williams-Ashman HG, Canellakis ZN. Polyamines in mammalian biology and medicine. *Perspect Biol Med.* 1979;22: 421–53.
47. Wu G, Morris SM. Arginine metabolism: Nitric oxide and beyond. *Biochem J.* 1998;336:1–17.
48. Morris SM. Arginine metabolism: Boundaries of our knowledge. *J Nutr.* 2007;137(1):602–9.
49. Kelly BL, Ferreira A. β -amyloid-induced dynamin 1 degradation is mediated by N-methyl-D-aspartate receptors in hippocampal neurons. *Journal of Biological Chemistry.* 2006;281(38):28079-28089.
50. Kabogo D, Rauw G, Amritraj A, Baker G, Kar S. β -amyloid-related peptides potentiate K⁺-evoked glutamate release from adult rat hippocampal slices. *Neurobiology of Aging.* 2010;31(7):1164-1172.
51. Texidó L, Martín-Satué M, Alberdi E, Solsona C, Matute C. Amyloid β peptide oligomers directly activate NMDA receptors. *Cell Calcium.* 2011;49(3):184-190.
52. Auguet M, Viossat I, Marin JG, Chabrier PE. Selective inhibition of inducible nitric oxide synthase by agmatine. *The Japanese Journal of Pharmacology.* 1995;69(3):285-287.
53. Satriano J, Schwartz D, Ishizuka S, Lortie MJ, Thomson SC, Gabbai F, Kelly CJ, Blantz RC. Suppression of inducible nitric oxide generation by agmatine aldehyde: Beneficial effects in sepsis. *Journal of Cellular Physiology.* 2001;188(3):313-320.
54. Mun CH, Lee WT, Park KA, Lee JE. Regulation of endothelial nitric oxide synthase by agmatine after transient global cerebral ischemia in rat brain. *Anatomy & Cell Biology.* 2010;43(3):230-240.
55. Ahn SK, Hong S, Park YM, Choi JY, Lee WT, Park KA, Lee JE. Protective effects of agmatine on lipopolysaccharide-injured microglia and inducible nitric oxide synthase activity. *Life Sciences.* 2012; 91(25):1345-1350.

© 2018 Çelik et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<http://www.sciencedomain.org/review-history/25606>